

## **Total RNA/mRNA Isolation**

**Reagents:** RNA STAT-60: red solution (TEL-TEST "B", INC, Friendswood, TX)  
Choloroform, Isopropanol, Ethanol

### **Procedure:**

#### **1. Homogenization:**

**Attached cells:** Add 1ml/35mm dish RNA STAT-60 directly into culture dishes and pass the cell lysate several times through a pipette. (DONOT WASH THE CELLS)

**Non-adherent cells:** cells are sedimented then lysed in RNA STAT-60 (1ml/5-10 x 10<sup>6</sup> cells) by repetitively pipetting.

- a. Add 0.2 ml of chloroform, cover the samples tightly, shake vigorously for 15 seconds, and then let samples stay at room temperature for 2-3 minutes.
- b. Centrifuge samples at 12,000xg for 15minutes at 4<sup>0</sup>C, and then transfer the aqueous phase (up clear lay) to a new tube.

#### **2. RNA precipitation**

- a. Add 0.5 ml of isopropanol into the new sample tube with aqueous phase, vortex to mix
- b. Store samples at room temperature for 5-10 minutes.
- c. Centrifuge at 12,000xg for 10minutes at 4<sup>0</sup>C. RNA precipitate forms a white pellet at the bottom of the tubes
- d. Remove supernatant

#### **3. RNA wash**

- a. Add 1ml cold 75% ethanol into the sample tubes, vortex, and then centrifuge for at 12,000xg for 5 minutes at 4<sup>0</sup>C.
- b. Pour the supernatant, dry the RNA pellet by lyophilizing for 3-5 minutes (DO NOT DRY COMPLETELY)
- c. Add 15ul DEPC water and pass the pellet through a pipette tip to dissolve the RNA pellet completely.

#### **4. Measurement of yield**

- a. Add 598ul of DEPC water into a new tube and add 2ul of RNA sample.
- b. Mix and measure OD at 260 nm (our machine reads: ug/ml)